



Mansoura University Faculty of Medicine Clinical Pathology Department

LOG BOOK

For Postgraduate Students
Applying for Doctor Degree in Clinical Pathology

Updated Version



رئيس القسم ا<u>.</u>د صلاح أغا

إعداد ا<u>.</u>د طارق سليم

Table of contents

Subject	Page number
Personal data	2
Policy of the log book	3
First part course	4-10
Second part compulsory courses	11-42
Second part elective courses	43-46
Scientific activities	47-51
Thesis	52
Appendix	53

Personal Data		
- Name:		
- Telephone:		
- E. mail address:		
- Date of graduation:		
- Degree:		
- Date of registration:		
- Date of Thesis appro	val:	
- Starting Date:		
Signature:		
Head of the Department	Vice Dean for research and postgraduat	e study

Policy of the log Book

Introduction:

According to the new postgraduate study regulation, postgraduate students applying for Master Degree in Clinical Pathology should fulfill a number of credit hours to be awarded. This is performed through attendance of scientific lectures, practical sessions as well as scientific activities including, thesis discussion, conferences and workshops in different specialties of Clinical Pathology. A ending at least 70% of different activities is a necessary requirement to get the permission to join the final degree exam.

Sections of the log book:

I-First part courses

II- Second part compulsory courses

III- Second part elective courses

IV- Scientific activities

V- Thesis

VI- Appendix

Assessment of the log book

Student's attendance and performance will be assessed by supervisors in different specialties of clinical pathology. Every activity or assessment should be documented by writing. The examination committee will revise the log book at the end of training before the final exam

I-First Part Course

Title of the course: Basics of Clinical Pathology

Course code: CPATH 630 BCP

Credit hours: 5

Teaching hours: 75

Scientific Lectures (Hematology)

Date	Title	Supervisor's signature
	The structure of the marrow and the hematopoil microenvironment	
	The lymphoid tissues	
	Hematopoietic stem cells.	
	Erythropoiesis	
	Granulopoiesis	
	Lymphopoiesis	
	Thrombopoiesis	
	Red cell structure and metabolism	
	Globin gene expression	
	Hemoglobin structure-function relationship	
	Iron homeostasis: Molecular control	
	Phagocytes	
	Physiology of hemostasis	
	Cell cycle regulation and apoptosis	
	Cellular signal transduction pathways	
	Principles of immunohematology	
	Quality assurance in hematology laboratory	
	Genetic principles and molecular biology	
	Cluster of differentiation antigens	
	Organization and management of Hematology laboratory	
	Automation in Hematology Laboratory	
	Hematology in under-resourced laboratories	

Scientific Lectures (Clinical Chemistry)

Date	Title	Supervisor's signature
	General lab. Supplies:	
	Lab. Glass ware	
	Types	
	Cleaning	
	Lab. plastic ware	
	Types	
	Cleaning	
	Volumetric equipments	
	- Pipettes (Method of calibration)	
	- Volumetric flasks	
	- Graduated cylinder	
	Laboratory operations	
	- Counting actions (Types /operation and	
	naintenance)	
	- weighing (Types / operation and maintenance)	
	Calculation in clinical chemistry	
	- Preparation of solution	
	- Dilution of concentrated solutions	
	- Percent concentrations	
	- Normal and Molar solutions	
	Specimen collection and handling	
	Collection	
	Types of samples	
	Preservation and transport	
	Separation and storage	
	Saliva an alternative to laboratory samples	
	Basic lab. Skills	
	- Units	
	- Centrifuges	
	- Balances	
	- Water	
	- Chemicals	
	- Reagent preparations	
	- Desiccants	
	- Mixers and homogenizers	
	- Pipettes	
	Spectrophotomety	
	- Types	
	- Components	

- Performance and standardization
Nephlometry and turbidmetry:
Principles and applications
Fluorometry.
- Principles and applications
Electrophoresis
-Types
- Factor affecting performance & results
-Technical considerations,
- Staining & clinical applications.
- Scanning
Chemiluminescence
Prilnciples and applications
Osmometry
- Types
- Clinical significance
Electrochemical techniques
- Potentiometry types of electrodes
- Amperometry principles and application (ISE)
- Biosensores
Chromatography
- Types
- Mechanism of separation
- HPLC & GC/MS
- Precautions
Qualitative (imunoelectrophoresis, gel diffusion)
- Qualitative
- RID
- Labeled assays (RIA, EIA)
Nanotechnology
Automation & dry chemistry
- Selection of an automated instrument
Point of care testing& Panic values
Q.C for Selection and evaluation of methods
Q.C for Selection and evaluation of methods
Quality management
- Control sample
- Calibrator
- Standard
- Types of Q.C
- Charts used for Q.C study e.g. L.J chart,
Westgard rolesetc
- Six sigma
- SDI
- Recovery and interferences
- Accuracy and precision
- Yes or no decision for laboratory run

Reference interval theory
- Basis
- Use
- Calculations
- Precautions during interpretation
- Normal range Vs reference interval
Screening tests
- Value
- Uses
- Disadvantages and how to overcome
Types & sources of error
- Technical
- Clerical
Body fluids methods of assay and evaluations
Molecular techniques in clinical chemistry, e.g.
PCR, FISH,
Proteomics.

Scientific Lectures (Clinical Microbiology)

Date	Title	Supervisor's signature
	Classification of microbes	
	Specimen collection and processing	
	Isolating media, tissue culture and media preparation	
	Quality control in clinical microbiology and biohazards	
	Automation and computer in clinical microbiology	
	Identification techniques and Stains in microbiology	
	Basics in diagnostic virology , Mycology , Non	
	conventional pathogen	
	Phenotypic and genotypic testing of micro-organisms	
	Antimicrobials : Principle of action & antimicrobial	
	policy and resistance	
	Microbial evolution	
	Guidelines for hospital acquired infections	
	Basic immune response to microorganism	
	Sterilization And disinfection	
	Quantification in microbiology	

Scientific Lectures (Clinical Immunology)

Date	Title	Supervisor's signature
	Immune response	
	Innate immunity & toll like receptors	
	B lymphocytes & Ig structure & diversity	
	T lymphocyte & TCR diversity & T reg	
	Complement system	
	MHC & Ag processing	
	NK cells & KIRs	
	Cytokines & chemokines	
	Tolerance & Autoimmunity	
	Molecular cell biology	
	Tests for immune competence	

II- Second Part Compulsory Courses

1- Hematology

Title of the course: Hematology

Course code: CPATH 630 HE

CPATH 630 HEP

Credit hours:

CPATH 630 HE: 8 hours

CPATH 630 HEP: 4 hours

Teaching hours:

CPATH 630 HE: 120 hours

CPATH 630 HEP: 120 hours

SCIENTIFIC LECTURES

Date	Title	Supervisor's signature
	General aspects of anemia	
	Iron-deficiency anemia	
	Iron refractory iron deficiency anemia	
	Iron overload	
	Anemia of chronic disease	
	Sideroblastic anemia	
	Hematological aspects of porphyria	
	Phenotype diversity of thalassemia and sickle cell anemia	
	Pre-implantation and prenatal diagnosis of thalassemia syndrome	
	Molecular techniques used in diagnosis of thalssemia syndrome	
	Molecular basis of thalassemia	
	Macrocytic anemias	
	General aspects of hemolytic anemias	
	Red cell membranopathies	
	Red cell enzymopathies	
	Hemoglobinopathies	

 T	T
Immune hemolytic anemias	
Non-immune hemolytic anemias	
Paroxysmal nocturnal hemoglobinuria	
Microangiopathic hemolytic anemias	
Aplastic anemia and Pure red cell aplasia	
Congenital dyserthropoietic anemias	
Pancytopenia	
Stem cell transplantation	
Benign disorders of granulocytes	
Benign disorders of lymphocytes	
Mononucleosis syndrome	
Benign disorders of monocytes	
Disorders of macrophages	
Hypersplenism and hyposplenism	
Oncogenesis	
Cancer stem cell	
Cytogenetics of hematological malignancies	
Molecular genetics of hematological malignancies	
Acute lymphoblastic leukemia	
Acute myeloid leukemia	
Myelodysplasia	
Chronic myeloid leukemia	
Polycythemia	
Myelofibrosis	
Primary thrombocythemia	

Chronic lymphocytic leukemias
Hodgkin`s lymphoma
Non Hodgkin`s lymphoma
Multiple myeloma
Essential monoclonal gammopathy
Macroglobulinemia
Heavy chain disease
Free immunoglobulin light chain
Amyloidosis
Angiogenesis
Vascular purpuras
Thrombocytopenia
Thrombocytosis
Hereditary qualitative platelet disorders
Acquired qualitative platelet disorders
Hemophilias
Molecular genetics of hemophilia
vonWillebrand's disease
Acquired coagulopathies
Circulating inhibitors of coagulation
Hereditary thrombophilia
Molecular genetics of thrombophilia
Acquired thrombophilia
Antiphospholipid syndrome
Thrombotic microangiopathies

Antithrombotic therapy	
Red cell antigens and antibodies	
Leukocytes and platelet antigen and antibodies	
Blood components therapy	
Complications of blood transfusion	
Autologous blood transfusion	
Hematologic aspects of systemic diseases	
Therapeutic apharesis	
Blood alternatives	
Hematology in pregnancy	
Neonatal hematology	
Geriatric hematology	

II- Practical Sessions

Skill	Level of performance			Trainee's assessment			Trainer's
	Observation	Assistance	Independence	Poor	Fair	Good	signature
Collection and handling of blood samples							
Tests for acute phase response ESR							
Plasma Viscosity Whole blood viscosity							
Hemoglobinometry							
Manual red cell count reticulocyte count, hematocrit and calculation of red cell indices							
Manual p Manual total and differential count latelet count,							
Automated blood count							
Preparation and staining methods of blood and bone marrow films							
Blood cell morphology in health and disease							

Examination of blood		
films parasites		
l mile paraertee		
Bone marrow		
aspiration		
Bone marrow		
Trephine biopsy		
<u>Laboratory</u>		
assessment of iron		
<u>status</u>		
Estimation of serum		
iron		
Estimation of total		
iron-binding capacity		
Estimation of serum		
ferritin		
Estimation of serum		
transferrin, transferrin		
saturation and index		
and transferrin		
receptors		
Assay of serum		
hepcidin		
Estimation of free		
erythrocyte		
protoporphyrin		
Laboratory toota		
<u>Laboratory tests</u> used in		
investigation of		
<u>megaloblastic</u>		
<u>anemia</u>		
Me Measurement of		
red cell folate		
asurement of serum		
B12		
Measurement of		
intrinsic factor		
antibody		
Schilling test		
Johnning test		
Measurement of		
serum gastrin		
Measurement of		
gastric juice PH		
gasaro jaroe i ii		

		,
Measurement of gastric juice pH		
Measurement of serum		
methylmalonic acid		
Measurement of serum homocysteine		
Measurement of serum transcobolamin		
Measurement of serum holotranscobolamin		
Generic laboratory tests used in the investigation of hemolytic anemias		
Measurement of seum bilirubin, haptoglobin and hemopexin		
Measurement of plasma hemoglobin, methemalbumin, methemoglobin, sulphemogloin and carboxyhemoglobin		
Demonstration of urobilinogen, hemosiderin, myoglobin and porphobilinogen urine		
Investigations of hereditary hemolytic anemias		
Erythrocyte membrane protein analysis		
Osmotic fragility test		
Glycerol lysis-time, Cryohemolysis and		
autohemolysis tests		
Methemoglobin reduction and fluorescent		
screening tests for		

G6PD deficiency		
Quantitative G6PD assay		
Detection of heterozygotes for G6PD deficiency		
Pyrimidine-5- nucleotidase screening test		
Identification of G6PD variants		
Pyruvate kinase assay		
Estimation of reduced glutathione		
Glutathione stability test		
Measurement of red cell 2,3-diphosphoglycerate		
Determination of the oxygen dissociation curve		
Investigation of hemoglobinopathies		
Detection of hemoglobin variants:		
Cellulose acetate electrophoresis at alkaline pH		
Citrate agar electrophoresis at pH 6		
Agarose gel electrophoresis		
Automated HPLC		
Isoelectric focusing		
Detection of unstable hemoglobins		
Detection of		

hemoglobin Ms		
Detection of altered		
affinity hemoglobins		
Sickling in whole		
blood		
biood		
HbS solubility test		
Investigation of		
<u>thalassemia</u>		
Quantitation of Hb F		
O		
Quantitation of Hb A2		
Assessment of the		
intracellular		
distribution of Hb F		
Fetal diagnosis of		
globin chain disorders		
Investigations of		
Investigations of acquired hemolytic		
anemias		
Antiglobulin		
(Coombs') test		
Acidified serum		
(Ham test)and		
Sucrose lysis tests		
Investigations of		
<u>hemostasis</u>		
Measurement of		
prothrombin time		
Measurement of		
activated		
thromboplastin time		
Measurement of		
fibrinogen		
concentration (Clauss		
method)		
Estimation of bleeding		
time		
Clot solubility test for		
FXIII		
Detection of fibrin		
L		

(ogen) split products and D-dimer		
Detection of circulating coagulation inhibitor		
Bioassays of coagulation factors		
Von Willebrand factor antigen assay		
Platelet aggregometry		
Detection of carriers of congenital coagulation deficiency defects		
Investigation of thrombotic		
<u>tendency</u>		
Clot-based assay for activated protein C resistance		
Detection of factor V		
Leiden and prothrombinG20210A mutations		
Antithrombin, protein C and protein S assays		
Euglobuin lysis test		
tPA, PAI-1 and α2- antiplasmin assay		
Markers of platelet and coagulation activation		
Global tests of coagulation		
Global protein c assay		
Thromboelastograph		
Laboratory aspects of transfusion medicine		

5 ()		
Pre-transfusion compatibility systems		
ABO and D grouping		
Antibody screening		
Antibody identification		
Selection and transfusion of red cells		
Cross-matching		
Compatibility testing in special transfusion situations		
Investigation of a transfusion reaction		
Erythrocyte cytochemistry		
Staining of siderotic granules		
Demonstration of Heinz bodies		
Demonstration of Hb H inclusions		
Demonstration of cellular Hb F (Kleihauer test)		
Leucocyte cytochemistry		
Myeloperoxidase		
Sudan black B		
Neutrophil alkaline phosphatase		
Acid phosphatase reaction		
Periodic acid Schiff		
Naphthol AS-D chloracetate esterase		
α-naphthyl butyrate		

esterase		
α -naphthyl acetate esterase		
Toludine blue stain		
<u>Immunophenotyping</u>		
Flow cytometry		
Immuncytochemistry		
<u>Diagnostic radioisotopes in</u> <u>hematology</u>		
Measurement of blood volume		
Splenic red cell volume		
Ferrokinetics		
Estimation of the life span of red cells in vivo		
Compatibility test		
Visualization of the spleen by scintillation		
Measurement of blood loss from GIT		
Measurement of platelet life span		
Flow Cytometry analysis in hematology		
Cytogenetic analysis in hematology		
Molecular analysis in		
hematology		
Interpretation of hematology laboratory		
<u>data</u>		
Reference ranges and		
normal values		
Analysis of instrumental data output		
Case studies in hematology		

2-Clinical Chemistry

Title of the course: Clinical Chemistry

Course code: CPATH 630 CC

CPATH 630 CCP

Credit hours:

CPATH 630 CC: 8 hours

CPATH 630 CCP: 4 hours

Teaching hours:

CPATH 630 CC: 120 hours

CPATH 630 CCP: 120 hours

N.B: In each subject; an introductory hint of the basis will be followed by updates in that subject in relation to management of the related clinical situation.

SCIENTIFIC LECTURES

Date	Title	Supervisor's signature
	Carbohydrate homeostasis	
	DM pathogenesis, C/P complications & diagnosis (Updates)	
	Classification of lipids & lipid metabolism	
	Cardiovascular risk factors	
	Metabolic syndrome	
	Amino acids classification and metabolism	
	Protein structure & metabolism	
	Acute phase proteins	
	Inborn error of metabolism I	
	Inborn error of metabolism II	
	Inborn error of fatty acids and organic acids metabolism	
	Physiology of normal renal functions & Glomeruler & tubular function tests	
	Chemical pathology of renal disorders	
	Water homeostasis & related factors	
	Electrolyte balance, electrolyte disturbance and there assay	
	Acid base balance disorders	
	Physiology of liver function & Liver function tests	

Chemical pathology of hepatic disorders	
Gastric function tests and gastric diseases	
Exocrine pancreatic function tests & pancreatic diseases	
Intestinal function tests & malabsorbtion syndromes	
Cardiac function study	
Diagnosis of ischemic heart diseases	
Rule of laboratory in diagnosis & follow up of heart failure & hypertension	
Clinical enzymology I	
Clinical enzymology II	
Clinical enzymology III	
Ca homeostasis & assay	
Phosphorous & Mg disorders & assay	
Markesr of bone turnover	
Vitamin assessment	
Multiple endocrine neoplasm	
Trace element assessment	
Nutrition and obesity	
Biochemical Tumor markers	
Hypothalamopituitary unit	
Hypothalamopituitary adrenal axis	
Hypothalamopituitary thyroid axis	
Pancreatic hormones	
Reproductive related disorders	
Clinical chemistry of pregnancy & fetal	

monitoring
Assessment of porphyrins and disorders of porphyrin metabolism
Iron homeostasis
Clinical chemistry of pediatric
Clinical chemistry of geriatric
Adipose tissue as an endocrine organ
Applications of molecular biology in clinical chemistry
Microarray in clinical chemistry
Therapeutic drug monitoring
Updates in Clinical Chemistry
 Genetic updates in clinical chemistry Metabolic updates in clinical chemistry Free radicals & oxidative stress Cytokines, endothelial markers & oxidant stress. Trace elements, elicit substance abuse & toxic elements in clinical chemistry field. Newly advanced markers in disease management & prognosis. Proteomics/genomics applications in clinical chemistry Nano-concepts in clinical lab. medicine

Practical Sessions

Skill	Level of performance			Trainee's assessment			Trainer's
	Observation	Assistance	Independence	Poor	Fair	Good	signature
Basic Lab supplies • Units. • Water. • Solutions Spectrometry							
Separation tech • Chromatography • Electrophoresis • GC/MS Osmometry							
 Potentiometry Amperometry POCT							
Q.C • Charts • Evaluations Reference values							
Immunoassays & automation							
Dry chemistry & nanotechnology							
Carobhydrates Glucose Specimen Andyticol method Ref. Value Oral glucose tolerance							

Chroate	od Uh	
Glycated Hb		
•	Specimen	
•	Analytical method	
•	Ref. Interval	
•	Clinical significance	
Lipog		
•	specimen	
•	analytical methods	
•	clinical significance	
•	LDL calculation	
•	Metabolic syndrome Risk assessment	
Proteir		
11000	•	
•	Plasma proteins and	
	albumin	
•	Specimen	
•	Analytical methods	
•	Clinical significance	
•	Ref. Values	
Urinary	/ protein	
	Specimen collection	
•	Analytical methods	
•	Clinical significance	
•	Ref. interval	
•	ACR (Albumin / create	
	ratio) `	
Non Pr	otein nitrogenous	
compo	und urea/ creatinine /	
uric aci	d.	
•	clinical significance	
•	analytical methods	
•	critical values	
•	specimen	
•	Ref. interval	
Electro	olytes	
•	- Na+, K+, CI-, HCO3-	
	- Clinical significance	
	_	
	- Specimen	
	- Analytical method	
	- Ref. interval	
	- Critical values	
	- Anion gap, osmal gap	

Blood gases		
Specimen		
Clinical significanceRef. Values		
Critical values		
Bone minerals: Ca, ph, Mg++		
Specimen		
 Clinical significance 		
Analytical methods		
Ref value Critical value		
Enzymes		
, .		
Liver Enzymes(ALT. AST, ALP,		
s'nucleotide& GT)		
Cardiac and skeletal		
ms (CK , LDH) • Pancreatic enzymes		
(amylase , lipase)		
Miscellaneous		
enzymes		
Urine analysis:		
Physical		
Chemical		
Seminal fluid analysis		
 CSF analysis 		
PhysicalChemical		
Peritoneal, pleural, synovial		
fluid and pericar dial fluid		
analysis.		
,		
Physical		
ChemicalMicroscopic		
Specimen		
Amniotic fluid testing:		
Cytogentic studiesAFP		
Acetyl cholin tore		
 Evaluation of fetal 		
lung maturity		
Tests for isoimmunization		
Evidence-based laboratory		
medicine		

3-Clinical Microbiology and Immunology

Title of the course: Clinical Microbiology and Immunology

Course code: CPATH 630 CMI

CPATH 630 CMIP

Credit hours:

CPATH 630 CM: 8hours

CPATH 630 CMP: 4 hours

Teaching hours:

CPATH 630 CM: 120 hours

CPATH 630 CMP: 120 hours

SCIENTIFIC LECTURES(Clinical Microbiology)

Date	Title	Supervisor's signature
	1- Introduction to clinical	
	microbiology	
	Cell structure, physiology, metabolism and genetics	
	Classification of microbes	
	Introduction to viral infections	
	Introduction to fungal infections	
	Bacterial flora	
	Performance improvement in the Microbiology laboratory	
	 Antimicrobial drugs Classification Mechanism of action 	
	Molecular diagnosis	
	Immunodiagnosis of infective syndromes	
	Disinfection and sterilization	
	Automation : principles of instruments used in microbiology lab.	
	* Lab. Identification of significant isolates :-	
	Staphylococci.streptococci, enterococci and other catalase positive Gram positive cocci	

Neisseria , Moraxella, Haemophilus and other	
fastidious gram negative Bacteria	
iasaalous grain negauve bacteria	
Enterobacteracae	
Vibria saramanas campulabactor and and	
Vibrio , aeromonas, campylobacter and and Non fermentive Gram negative bacilli	
Non Termentive Grain negative bacilli	
Anaerobes	
Chlamydia & Rickettsia	
Mycoplasma and ureaplasma	
mycopiasina anu ureapiasina	
3- Lab. diagnosis of infectious	
diseases :	
Upper and lower respiratory tract infections	
Skin and soft tissue infections	
Anaerobic infections	
GIT and food poisoning	
Infection of central nervous system	
Bacteraemia and septicaemia	
Huinam August infrastions	
Urinary tract infections	
Genital infections and sexually transmitted	
diseases	
Infection in special population :	
- In transplant patients	
- In children	
- In HIV patients	
- Opportunistic infection	
Occular infections	
Pyrexia of unknown origin	
Mycobacterial infection	

Classification	
- Classification	
- Pathophysiology	
- Epidemiology	
- Multidrug resistance & its	
mechanism	
- Public health concern	
4-Clinical syndromes associated	
with viral infections	
5- Fungal infections	
- Pathophysiology	
- Transmission	
- Clinical presentations	
- Epidemiology	
- Diagnosis (Superficial & deep) .	
Infection control	
Parasitic infections	
- Intestinal parasites	
- Tissue parasites	
- Blood parasites	
- Protozoa	
* Biofilm	
* Host – parasite interrelationship	
* Antimicrobial drug resistance &mobile genetic elements	
* Probiotics	
* Bioterrorism	
* Biohazard and Biosafety	
24	

Air and water pollution	
- Testing of quality	
- related Biohazards	
* Nanotechnology in clinical microbiology	
* Emerging pathogens	
* Infection control guidelines	
* Public health principles and interrelation between diagnostic lab. and public health agencies	
* Chronic fatigue syndrome (Microbiological causes)	

Practical Sessions(Clinical Microbiolgy)

Skill	Level of perfo	ormance		Trainee'	's assessm	ent	Trainer's
							signature
	Observation	Assistance	Independence	Poor	Fair	Good	
Specimen collection in					•	•	
the microbiology lab.							
Microscopes in microbiological diagnosis							
Media used for isolation							
of microorganisms							
Staining techniques and							
its interpretation							
Presumptive identification							
of bacterial growth :-							
• Colonial							
morphology							
 Biochemical identification(man 							
ual & automated)							
 Antimicrobial susceptibility 							
tests							
 principle of (MICs, breakpoints, disc 							
diffusion, agar							
dilution).							
Immunodiagnosis of							
infective syndromes							
Automation							
Application of Molecular							
methods in diagnostic							
microbiology							
• DNA & RNA							
extraction							
AmplificationDetection (gel							
electrophoresis)							

Schematic outlines of		
microbiological diagnosis		
Examination of :-		
Sputum Throat & mouth specimens Pus, ulcer material, skin specimens Effusion C.S.F Urine Stool Urogenital sample & semen Blood		
Sterilization method		
Anaerobic isolation & identification		
Water related dis. and		
testing of water supplies		
Virology		
SpecimenTransportMethods		
Mycology		
 Specimen Direct examination Culture Interpretation of results 		
ТВ		
 Staining methods of Acid fast bacilli Interpretation of ZN stained smear Decontamination procedures Methods of Mycobacterial culture & identification Molecular 		

methods for diagnosis of Mycobacteria Indirect methods of diagnosis of Tuberculosis Antituberculous susceptibility testing	
Interpretation of	
microbiological results	
Atypical organisms	
MycoplasmaLeigonellaChlamydia	
Spirochates	

SCIENTIFIC LECTURES (Clinical Immunology)

Date	Title	Supervisor's signature
	Introduction to Immune System	
	Immune response part I	
	Immune response part II	
	Innate Immunity Part I	
	Innate Immunity Part II	
	Antigen and Immunogene	
	T-lymphocytes & Tregs	
	B-lymphocytes	
	Immunoglobulin	
	Receptor Diversity	
	NK and KIR	
	Antigen-presenting cell , Ag processing and presentation	
	Complement system	
	Major histocompatibility complex	
	Histocompatibility testing	
	Cytokines	
	Chemokines	
	Tolerance & Autoimmunity	
	Evaluation of Immune-competence	
	Immunodeficiency syndromes Part I	
	Immunodeficiency syndromes Part II	
	Hypersensitivity Part I	
	Hypersensitivity Part II	
	Immune-mediated Rheumatic diseases Part I	

Immune-mediated Rheumatic diseases Part II
Immune-mediated GIT& Hepatobiliary diseases
Immune-mediated Endocrine dis.
Tumor Immunology
Transplant Immunology Part I
Transplant Immunology Part II
Stem cell transplant
Immunologic therapy
Reproduction and immune system
Immune-mediated hematologic diseases
Inflammation
Mucosal Immunity
Immune-mediated vascular disease
Immune-mediated neurologic disease
Immune modulating therapy
Gene therapy
Cell cycle kinetics
Introduction to molecular biology
Molecular biology in immunology

Practical Sessions(Clinical Immunology)

Skill	Level of perfo	ormance		Traine	e's asse	essment	Trainer's
	Observation	Assistance	Independence	Poor	Fair	Good	signature
				1001	lan	dood	
Immunodiffusion methods							
Nephlometry							
Immunoelectrophoresis							
methods							
Enzyme immunoassay							
Immunoflourescence methods							
Agglutination assays							
Complement assays							
Lymphocyte separation							
Lymphocyte activation							
Flowcytometery principles and applications							
Lymphocyte assays							
Neutrophil function							
HLA-typing by serology							
Cross match							
Cellular typing & PRA							

PCR principles and		
applications		
How to prepare solution for		
molecular biology		
DNA extraction		
HLA-typing by Innolipa		
Agarose gel elctrophoresis		
HLA-typing by SSP		
RFLP		
KFLP		
ARMS		
ANIIIO		
Allergy & Immunocap		
,		
Instrument in cl.		
Immunology lab		
<u> </u>		

III- Second Part Elective Courses

1- Stem Cells

Title of the course: Stem Cells

Course code: CPATH 630 SC

Credit hours: 1

Teaching hours: 15

Scientific Lectures

Date	Title	Supervisor's signature
	Stem cell: General issues	
	Stem cell separation	
	Stem cell culture	
	Stem cell niche	
	Immunogenetics of stem cells	
	Cancer stem cell and leukemic stem cell	
	Stem cell transplantation	
	Stem cell plasticity	

2- Principles of Molecular Genetics

Title of the course: Principles of Molecular Genetics

Course code: CPATH 630 PMG

Credit hours: 1

Teaching hours: 15

Scientific Lectures

Date	Title	Supervisor's signature
	Basic DNA structure	
	Gene structure and expression	
	Methods of DNA analysis	
	Gene mutations	
	Gene polymorphisms	
	The cell cycle	
	Molecular basis of Cancer	
	DNA-based HLA typing	
	Microbial genetics	
	Microbial phylogenetic analysis	
	Human Genome project, genomics & epigenetics	

IV- Scientific activities

1-Thesis discussion attendance:

Date	Title of the thesis	Supervisor's signature

2-Conference attendance:

Date	Title of the Conference	President's Signature

3-Workshop attendance:

Date	Workshop title	Organizer' s signature

V- Thesis

Credit hours: 15

Title of the thesis	Date of discussion	Principal supervisor's signature

VI-Appendix